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Thin-layer chromatography of inorganic sulphur compounds

Procedures have been described for the separation of inorganic sulphur compounds by paper¹⁻⁵ and ion-exchange⁶⁻⁸ chromatography, but few systems using thin-layer chromatography have been described^{9,10}. Owing to the high reactivity of thiosulphate and polythionates in the mixtures encountered in metabolic studies of such compounds^{1,7,11}, a method for the rapid analysis of these mixtures was desirable. The procedures described below enable separations of sulphate, thiosulphate, trithionate, tetrathionate and thiocyanate to be made.

Experimental

Stationary phases were sheets of Gelman instant thin-layer chromatography (ITLC) media, types SA and SG (20 × 20 cm). Chromatograms were run in Gelman ITLC chambers at 22–25°. Marker compounds (usually 10 µg in 10 µl water) were applied 3 cm from the edge of the sheets and the solvent allowed to run 13 cm beyond the origin line. Sources of marker compounds were as follows: AnalaR grade KSCN and K₂S₂O₃ from the British Drug Houses; ³⁵S-labelled sulphate and thiosulphate from The Radiochemical Centre (Amersham, Great Britain); ³⁵S-labelled thiocyanate was synthesised by the cyanolysis of labelled tetrathionate; and potassium tetrathionate¹² and trithionate¹³ were synthesised. Solvents were AnalaR or best commercial grades and were not further purified. Sulphur compounds were detected by spraying dried chromatograms with 8% (w/v) silver nitrate in acetone containing 10% (v/v) water, and by radioautography and liquid scintillation counting when ³⁵S-labelled materials were used.

Results and discussion

A number of solvents, including several suitable for paper chromatography, were generally unsatisfactory for TLC separations in this study (Table I). Solvent S₂ is very useful for paper chromatography², and did separate the four compounds tested. The rather large spots produced did, however, decrease the efficiency of sepa-

TABLE I

TLC OF THIONATES AND THIOCYANATE ON SA MEDIUM USING MIXED SOLVENTS

Solvents used were: S₁ 1-butanol–acetone–water (40:40:30)
 S₂ 1-butanol–pyridine–acetic acid–water (30:20:6:24)
 S₃ 1-butanol–pyridine–acetic acid–water (90:3:1:6)
 S₄ 1-propanol–pyridine–water (50:35:50)
 S₅ ethanol–pyridine–water (80:10:5)
 S₆ ethanol–pyridine–water (60:30:10)
 S₇ 2-heptanol–methanol–water (85:10:5)

Compound	<i>R_F</i>						
	S ₁	S ₂	S ₃	S ₄	S ₅	S ₆	S ₇
Thiosulphate	0.68	0.50	0.02	0.84	0.45	0.54	0.05
Trithionate	0.76	0.61	0.26	0.86	0.72	0.74	0.05
Tetrathionate	0.79	0.67	0.39	0.86	0.78	0.76	0.05
Thiocyanate	0.84	0.78	0.62	0.82	0.76	0.75	0.22

TABLE II

TLC OF INORGANIC SULPHUR COMPOUNDS ON SA MEDIUM USING ALCOHOLS AS SOLVENTS

Solvents used were: S₈ methanolS₉ 99% ethanolS₁₀ 5% (v/v) water in 1-propanolS₁₁ 5% (v/v) water in 1-butanolS₁₂ 5% (v/v) water in 1-pentanolS₁₃ 5% (v/v) water in 1-hexanolS₁₄ 1-octanol saturated with waterS₁₅ 1-propanol-methanol (1:1)S₁₆ 5% (v/v) water in 2-butanol

Compound	R_F								
	S ₈	S ₉	S ₁₀	S ₁₁	S ₁₂	S ₁₃	S ₁₄	S ₁₅	S ₁₆
Sulphate	0 ^a	0	0	0	—	—	—	0 ^a	—
Thiosulphate	0.78	0.25	0.06	0.02	0.10	0	0	0.49	—
Trithionate	0.84	0.73	0.54	0.35	0.09	0.03	0.03	0.73	0.27
Tetrathionate	0.84	0.80	0.63	0.46	0.19	0.10	0.04	0.77	0.37
Thiocyanate	0.83	0.79	0.71	0.64	0.47	0.38	0.36	0.77	0.60
Approx. time for standard run (min)	45	85	140	180	300	390	520	90	240

^a See text.

ration of thiocyanate and the polythionates from each other. Thiocyanate was separated effectively from all the other compounds only by solvent S₇.

In contrast, all four compounds and sulphate could be separated from each other using pure alcohols or alcohols containing small quantities of water (Table II). No single solvent effected perfect separation of all five compounds, but 5% (v/v) water in 1-butanol (S₁₁) gave complete separation of thiosulphate, trithionate, tetrathionate and thiocyanate, while methanol (S₈) or methanol-propanol (S₁₅) completely resolved sulphate from the other compounds. Solvent S₁₅ allowed isolation of sulphate, thiosulphate and thiocyanate or polythionates from each other. Resolution of a mixture of all five compounds can thus be achieved by two-dimensional chromatography in solvents S₁₅ and S₁₁. The behaviour of sulphate in solvents S₁₅ and S₈ indicated S₁₅ to be preferable. Sulphate showed no significant movement in ethanol or longer-chain alcohols, but tended to streak from the origin in solvents S₁₅ and S₈. This streaking was more pronounced with S₈ and the distance streaked increased with the amount of sulphate on the thin layer. When 30 μg of sodium sulphate was applied, 80% remained within 2 cm of the origin, but the remainder streaked a further 7 cm in S₈. With methanol-propanol (S₁₅), 80–85% of the sulphate remained within 2 cm of the origin, most not migrating at all, and none moved further than 3.5 cm from the origin. The "tail" of the sulphate thus remained 1.5–2 cm behind the rear of a 20 μg spot of potassium thiosulphate.

Pure 1-propanol was an unsatisfactory solvent, as sulphate and thiosulphate remained at the origin, while trithionate and tetrathionate gave spots of R_F 0.36 and 0.54 but streaked from the origin.

Gelman ITLC SG medium was not generally suitable for these separations, although with solvent S₁₄ thiocyanate (R_F 0.78) was separated from all the other compounds, which did not migrate.

In conclusion, chromatography on Gelman ITLC SA media using 5% water in 1-butanol and a 1:1 mixture (v/v) of methanol-1-propanol effects complete separation of sulphate, thiosulphate, trithionate, tetrathionate and thiocyanate in not more than 5 h. This technique is of use in analysing mixtures such as are encountered during bacterial sulphur metabolism^{14,15} or during reaction of thionates with cyanide¹⁶.

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- 1 P. A. TRUDINGER, *Aust. J. Biol. Sci.*, 17 (1964) 459.
- 2 P. A. TRUDINGER, *J. Bacteriol.*, 93 (1967) 550.
- 3 F. H. POLLARD, D. J. JONES AND G. NICKLESS, *J. Chromatog.*, 15 (1964) 393; F. H. POLLARD, G. NICKLESS, D. J. JONES AND R. B. GLOVER, *J. Chromatog.*, 15 (1964) 407.
- 4 B. SKARZYNSKI AND T. W. SZCZEPKOWSKI, *Nature*, 183 (1959) 1413.
- 5 M. OKUZUMI AND K. IMAI, *J. Ferment. Technol.*, 43 (1965) 10.
- 6 A. IGUCHI, *Bull. Chem. Soc. Japan*, 31 (1958) 597, 600.
- 7 P. A. TRUDINGER, *Aust. J. Biol. Sci.*, 17 (1964) 446.
- 8 F. H. POLLARD, G. NICKLESS AND R. B. GLOVER, *J. Chromatog.*, 15 (1964) 533.
- 9 H. SEILER AND H. ERLNMEYER, *Helv. Chim. Acta*, 47 (1964) 264.
- 10 F. H. POLLARD, G. NICKLESS, K. BURTON AND J. HUBBARD, *Microchem. J.*, 10 (1966) 131.
- 11 D. P. KELLY, *Aust. J. Sci.*, 31 (1968) 165.
- 12 P. A. TRUDINGER, *Biochem. J.*, 78 (1961) 680.
- 13 H. STAMM AND M. GOEHRING, *Z. Anorg. Allgem. Chem.*, 250 (1942) 226.
- 14 D. P. KELLY AND P. J. SYRETT, *Biochem. J.*, 98 (1966) 537.
- 15 P. A. TRUDINGER, *Advan. Microbiol. Physiol.*, 3 (1969) 111.
- 16 D. P. KELLY, L. A. CHAMBERS AND P. A. TRUDINGER, *Anal. Chem.*, 41 (1969) 898.

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